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In Silico Evaluation of *rbc*L, *mat*K, and *psb*A-*trn*H Regions on the Genus *Spatholobus* (Fabaceae)

Ahmad Muwaffiq Faza¹), Amira Khairunnisa Hidaya¹), Hafidza Fatma Yona¹), Twistka Talitha Pangestu¹), Muhammad Shafala Safa¹), Eko Suyanto^{1,2}), Turhadi^{1*}), Fatchiyah^{1,2})

¹)Biology Department, Faculty of Mathematics and Natural Sciences, Universitas Brawijaya. Jl. Veteran Malang, East Java, Indonesia, 65145

²)Research Center of Smart Molecule of Natural Genetics Resource, Universitas Brawijaya, Malang, East Java,

Indonesia, 65145

* Corresponding author

e-mail: turhadibiologi@ub.ac.id

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ABSTRACT

Spatholobus is a genus that belongs to the Fabaceae that is known to contain various bioactive compounds and distributed across Asia, including Indonesia. However, exploration of Spatholobus in Indonesia is still rare. Therefore, DNA barcoding is used to support the exploration and conservation of Spatholobus in Indonesia. However, there is no universal marker that can be used across all plant species. In addition, there are still few studies related to DNA barcoding within the genus Spatholobus. The purpose of this study was to evaluate the rbcL, matK, and intergenic spacer psbA-trnH regions in silico that can be used as DNA barcodes for the genus Spatholobus. This study began with a sequence search on the NCBI database including the rbcL, matK, and intergenic spacer psbA-trnH genes in the genus Spatholobus and Phaseolus coccineus as the outgroup. Each sequence was then aligned with ClustalW. Then, a phylogenetic tree was constructed using the Maximum-Likelihood (ML) with 1000× bootstrap. As a result, the rbcL, matK, and psbA-trnH regions can be used as markers for DNA barcoding in the genus Spatholobus with different specifications. The rbcL and matK can be used to distinguish Spatholobus at the genus level, while the psbA-trnH can be used to distinguish Spatholobus at the species level.

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INTRODUCTION

The genus *Spatholobus* is a plant from the Fabaceae family (Prasetyorini et al., 2022). This genus has been known as a traditional medicine that contains various bioactive compounds, such as terpenoids, alkaloids, flavonoids, and steroids. In addition, the content in this plant has been reported to have antioxidant, antibacterial, and anti-inflammatory activities (Rousdy et al., 2023; Yeni et al., 2023). Additionally, this plant has been widely studied as a plant that has anticancer activity (Hasna et al., 2022). This genus has 29 species distributed in the Asian region. Some of them, such as S. *suberectus* and S. *littoralis*, have been recognized

by the people of Borneo, especially the Dayak tribe as traditional medicines (Wardah & Sundari, 2019).

The genus Spatholobus is very diverse in Indonesia. However, the most recognized and widely studied species are still limited to S. suberectus and S. littoralis, also known as Bajakah wood. In fact, Spatholobus species includes more species in Indonesia, including S. albus, S. apoensis, S. auricomus, S. ferrugineus, S. gyrocarpus, S. hirsutus, S. latibractea, S. latistipulus, S. littoralis, S. macropterus, S. maingayi, S. multiflorus, S. oblongifolius, S. persicinus, S. ridleyi, S. sanguineus, and S. viridis. Most of these species are distributed on the island of Borneo, although some species are also distributed in Sumatra and Java (POWO, 2024).





The lack of exploration of the genus *Spatholobus* could lead to insufficient attention and conservation efforts on *Spatholobus* in Indonesia. Therefore, *Spatholobus* in Indonesia requires conservation efforts through a genetic approach. Genetic conservation is carried out to maintain the existence of a population by enriching existing genetic variation, because genetic diversity can determine the success of the population to adapt. Furthermore, genetic conservation through the identification process can help in the management of endangered species. One of these identification methods is through the DNA barcoding approach (Handayani & Setia, 2021; Willi et al., 2022).

DNA barcoding is an approach to identify organism or taxonomic classification using one or a few standard loci, so that the species can be distinguished from another. One commonly used barcode is the cytochrome oxidase 1 (CO1) mitochondrial gene, as a standard barcode for animal DNA barcoding. However, CO1 is not suitable for DNA barcoding in plants due to the low rate of nucleotide substitution. Therefore, finding a standard barcode for plants is quite challenging. Researchers have proposed various barcodes for plants, either from the nucleus such as ITS1 and ITS2, or the coding and non-coding regions of plastids such as rbcL, matK, rpoB, psbA-trnH, trnL-F,and others (Hollingsworth et al., 2011). The application of DNA barcoding in plants has been utilized in various fields, including for species identification (Nderitu et al., 2023), conservation of endangered and endemic plants (Rosario et al., 2019), and authentication of medicinal plants (Husin et al., 2018).

DNA barcoding as a strategy in *Spatholobus* conservation efforts has several challenges. First, there is currently no universal marker that can be used in all plant species (Li et al., 2014). In addition, there has been limited research on DNA barcoding of the genus *Spatholobus*. Therefore, it is necessary to evaluate markers in the plastid region in silico, especially *rbcL*, *mat*K, and intergenic spacer *psbA-trn*H as an initial step to study the genetic diversity of *Spatholobus* in Indonesia. In silico DNA barcoding is considered an effective and applicable approach to initiate various studies, such as taxonomic conservation of biodiversity (Pratiwi et al., 2023).

MATERIALS AND METHODS Data Mining

DNA sequences of the rbcL, matK, and psbAtrnH genes belonging to the genus Spatholobus and the outgroup Phaseolus coccineus were retrieved from the **NCBI** nucleotide database (URL: http://www.ncbi.nlm.nih.gov). All the sequences were critically evaluated and low-quality sequences (ambiguous base N > 3%) were discarded (Suesatpanit et al., 2017). The exclusion criteria used in this analysis were as follows: first, no whole genome data exceeding 150,000 bp in length were used (Su'udi et al., 2023). Second, no sequences from unclear species were used (Suesatpanit et al., 2017). Third, no sequences from mitochondria were used. Finally, no sequences that are too short were used (Sundari et al., 2022).

Data Analysis

The sequences obtained were then aligned with Clustal W in MEGA X software (Thompson et al., 1994; Kumar et al., 2018). The sequences were then compared with each other to determine the differences. In addition, the mutation pattern of the sequences was also analyzed. Then, a phylogenetic tree was constructed using the Maximum-Likelihood (ML) algorithm with a bootstrap of 1000×. The fittest model for ML was selected based on Bayesian Information Criterion (BIC) score (Nei & Kumar, 2000). The phylogenetic tree was constructed using MEGA X software (Kumar et al., 2018).

RESULT AND DISCUSSION

Data mining of Spatholobus sp. nucleotide sequences in the NCBI database was performed using various exclusion criteria. The first criterion is that whole genome sequences longer than 150,000 base pairs were not used. Barcoding using whole genomes with excessively long sequences requires significant time and high-performance computing to manage large whole genome data, making it ineffective (Su'udi et al., 2023). In contrast, sequences that are too short were also not used. Based on previous studies, molecular markers with sequence lengths above 500 bp are more effective in identifying plant species (Sundari et al., 2022). Furthermore, as the loci used were derived from chloroplasts, sequences derived from mitochondria were not included in the analysis. Based on the results of data mining using NCBI, several matK sequences were obtained from mitochondria, while matK is a maturase K protein-



coding gene found in plant chloroplasts (Kalangi et al., 2014). Therefore, matK sequence data from mitochondria were not used to avoid ambiguity in the results obtained. To ensure the precision of the data set, sequences with ambiguous species specifications, such as those that were only known as Spatholobus sp., were also excluded (Suesatpanit et al., 2017) (Table 1).

As previously outlined by Letsiou et al. (2024), barcoding studies of plant such as Spatholobus sp. generally use several specific loci derived from chloroplast that are known as genetic markers. Only four loci derived from chloroplast were found for the Spatholobus entry in NCBI, including rbcL, matK, psbA-trnH, and trnL-trnF. The data mining results showed that there are only 24 sequences for the *rbc*L, 14 sequences for the *mat*K, and 14 sequences for the *psb*A-*trn*H. Furthermore, sequences from the *trnL-trn*F locus were also found, but the species is unclear (KF621116.1 Spatholobus sp.). Therefore, trnL-trnF was not included in the analysis. As a result of the selection and exclusion process based on these criteria, the accession number used for analysis was 17 for rbcL, 8 for matK, and 14 for psbA-trnH (Table 1).

The Multiple Sequence Alignment (MSA) results on the *rbc*L sequences revealed 12 bases that differed among the species of S. suberectus, S. gyrocarpus, S. parviflorus, S. pulcher, and S. acuminatus. These differences are caused by point mutations. Point mutation is a specific type of mutation pattern that occurs in the DNA base sequence. According to Suprasanna et al. (2015), changes in the sequence of nitrogenous bases in DNA can lead to phenotypic changes (Suprasanna

et al., 2015). The alignment results of the *rbc*L gene demonstrated intraspecies base differences with a low frequency, indicating a relatively high level of homology. The 24th, 243rd, 300th, and 375th nucleotides (the 1st position is the first nucleotide after trimming and alignment), highlighted in red to illustrate divergences within the same species (intraspecies). The results of the rbcL alignment also showed base differences between different species (interspecies) (Table 2).

Additionally, slight base differences were observed at the *mat*K locus within the same species (intraspecies). A single base difference was noted in S. parviflorus at position 561 (Table 3). Mutations that occur at the matk locus are point mutations and insertion-deletion mutations (InDels). InDels, which are characterized by a gap (-), result from mismatches in the alignment of base sequences between species (Anzani et al., 2021). The results of the alignment using the *rbc*L and *mat*K indicate that these two loci are only applicable for Spatholobus studies at the genus level (interspecies).

The base sequence alignment in *Spatholobus* using the psbA-trnH locus identified the presence of InDel mutations and point mutations. Alignment results showed many base differences in nucleotide sequences within the same species (intraspecies), highlighted in red (Table 4). These results indicate that the psbA-trnH locus can be used for Spatholobus studies at the species level. Previous research has demonstrated the efficacy of the *psbA-trn*H locus for DNA barcoding in Stelechocarpus, indicating its potential as a DNA barcode at the species level (intraspecies) (Turhadi & Hakim, 2023)

Exclusion criteria	Num	ber of accession num	bers ⁽¹⁾
-	<i>rbc</i> L	matK	psbA-trnH
Whole genome	6	6	0
Unclear species	0	1	0
Mitochondrial sequence	0	8	0
Sequence that are too short	1	1	0

____ . .

Note: (1) Number of accession numbers that are not used





Sample					Nu	cleotid	le Sequ	lence				
Sumpre	24	81	96	168	169	197	203	243	300	303	306	375
KP313866.1 S. suberectus	G	G	Т	Т	С	G	Т	А	С	Т	С	Т
KP313865.1 S. suberectus	G	G	Т	Т	С	G	Т	А	С	Т	С	Т
KP313864.1 S. suberectus	G	G	Т	Т	С	G	Т	А	С	Т	С	Т
KP313863.1 S. suberectus	G	G	Т	Т	С	G	Т	А	С	Т	С	Т
KP313862.1 S. suberectus	G	G	Т	Т	С	G	Т	А	С	Т	С	Т
KP313861.1 S. suberectus	G	G	Т	Т	С	G	Т	А	С	Т	С	Т
KF181528.1 S. suberectus	А	G	Т	Т	С	G	Т	А	С	Т	С	С
KP202396.1 S. suberectus	G	G	Т	Т	С	G	Т	А	С	Т	С	Т
KP202395.1 S. suberectus	G	G	Т	Т	С	G	Т	А	С	Т	С	Т
KP202394.1 S. suberectus	G	G	Т	Т	С	G	Т	А	С	Т	С	Т
JF949991.1 S. suberectus	G	G	Т	Т	С	А	Т	А	С	Т	С	С
MG816933.1 S. gyrocarpus	G	G	С	С	Т	А	А	А	С	G	Т	С
MG816918.1 S. gyrocarpus	G	G	С	С	Т	А	А	G	С	G	Т	С
AB045825.1 S. parviflorus	G	G	Т	Т	А	А	Т	G	А	А	С	С
AB925446.1 S. parviflorus	G	G	Т	Т	А	А	Т	А	С	А	С	С
KF181563.1 <i>S. pulcher</i>	G	G	С	С	А	А	А	А	С	G	Т	С
AB925736.1 S. acuminatus	G	С	С	С	С	А	А	А	С	G	С	С

Table 2. Nucleotide differences in the rbcL locus of Spatholobus

*Red: intraspecies variation

Based on the MSA results, one of the factors that cause nucleotide sequence differences is transition and transversion (Monalisa & Lengkong, 2019). Transition itself is an exchange of bases between purines (A > G, G > A), or pyrimidines (C > T, T > C). Meanwhile, transversion is the exchange between purine bases and pyrimidine, or vice versa (A > C, A > T, G > C, G > T, C > A, C > G, T > A, T > G) (Hanifa et al., 2019). The overall mutation pattern showed a higher transition value compared to transversion. According to Rahayu et al (2019), transitions tend not to cause significant phenotypic changes. However, a higher transition value may indicate a mutation bias that could affect the accuracy of DNA barcodes. In the study conducted by Li et al. (2014), a low transition or transversion ratio was identified as a characteristic of a locus suitable for use as a DNA barcode in evolutionary studies.

Some nucleotide mutations can cause changes in the amino acids and the genomic sequence of chloroplast DNA. Changes in amino acids can affect the function of genes, thus changing their efficacy (Mahfut et al., 2019). Other research suggested that not all mutations can cause amino acid changes. Nonsense mutations can alter the genetic code that modify the amino acids produced. This can affect protein function due to changes in amino acid structure. Synonymous replacement mutations, on the other hand, only alter the genetic code without changing the amino acids produced. As a result, these mutations are usually neutral and do not significantly affect protein function because there is no change in amino acid structure, and they are not affected by natural selection (Sen et al., 2011).

The MSA results indicated that the conserved site in the *rbc*L mutation was 97.42%, while the variable site was 2.58%. The percentage of transitions in the *rbc*L mutation pattern was 0.44%, while the percentage of transversions was 0.22% (Table 5). The *psb*A-*trn*H mutation pattern showed that the conserved site at the *psb*A-*trn*H locus was 92.79%, while the percentage of variable sites was 7.21%.



													Nı	icleot	ide se	quenc	e									
Sample	2	5	6	21	30	36	40	55	59-61	79	85-89	109	133	142	145	161	163-167	198-201	206	217	220- 221	261	264- 267	270	272	299
GU396724.1 S. suberectus	А	Т	G	-	G	G	G	G	TTT	A	-	С	Т	А	Т	Т	-	ATAT	С	А	СТ	G	GAAA	G	Α	Т
GU396723.1 S. suberectus	А	Т	G	-	G	G	G	G	AAA	А	-	С	Т	А	Т	Т	-	ATAT	С	А	СТ	G	GAAA	G	А	Т
KX346984.2 S. suberectus	А	Т	G	-	А	Т	Т	G	TTT	A	-	С	G	С	Т	Т	-	A—T	С	A	СТ	G	GAAA	G	A	Т
KX346983.2 S. suberectus	А	Т	G	-	А	Т	Т	G	TTT	А	-	С	G	С	Т	Т	-	А—Т	С	А	СТ	G	GAAA	G	А	Т
MT077510.1 S. suberectus	Т	С	G	С	G	G	G	G	TTT	A	-	С	Т	А	Т	Т	-	ATAT	С	A	СТ	G	GAAA	G	A	Т
MT077509.1 S. suberectus	Т	С	G	С	G	G	G	G	TTT	A	-	С	Т	А	Т	Т	-	ATAT	С	А	СТ	G	GAAA	G	А	Т
MT077508.1 S. suberectus	Т	С	G	С	G	G	G	G	TTT	A	-	С	Т	А	Т	Т	-	ATAT	С	A	СТ	G	GAAA	G	A	Т
MT077507.1 S. suberectus	Т	С	G	С	G	G	G	G	TTT	A	-	С	Т	А	Т	Т	-	ATAT	С	A	СТ	G	GAAA	G	A	Т
MT077506.1 S. suberectus	А	Т	G	-	G	G	G	G	AAA	А	-	С	Т	A	Т	Т	-	ATAT	С	А	СТ	G	GAAA	G	А	Т
HG005078.1 S. suberectus	А	Т	G	-	А	Т	Т	G	TTT	A	-	С	G	С	Т	Т	-	A–T	С	A	СТ	G	GAAA	G	A	Т
MT077505.1 S. pulcher	А	Т	Т	-	G	G	G	С	AAA	А	CTTTT	А	G	С	G	А	TATGT	-	А	С	-	Т	-	А	Т	С
MT077504.1 S. pulcher	A	Т	Т	-	G	G	G	С	AAA	A	CTTTT	А	G	С	G	A	TATGT	-	А	С	-	Т	-	А	Т	С
MT077503.1 S. pulcher	A	Т	Т	-	G	G	G	С	AAA	А	CTTTT	А	G	С	G	А	TATGT	-	А	С	-	Т	-	А	Т	С
HG005110.1 S. pulcher	A	Т	Т	-	G	G	G	С	AAA	С	CTTTT	А	G	С	G	A	TATGT	-	А	С	-	Т	-	А	Т	С

Table 3. Nucleotide differences in the matK locus of Spatholobus

Table 4. Nucleotide differences in the *psbA-trn*H locus of *Spatholobus*

		Nucleotide Sequence																														
Sample	1 7	19-21	51-56	57	116	135	141	155	185	186	197	214	239	282	290	325	346	365	373	397-399	401-403	422	423-427	447	450	497	500	506	534	545	561	568
EU106113.1 Spatholobus parviflorus	G C	:	CTATTC	С	С	G	С	А	С	С	А	С	С	А	G	А	G	С	G	ACC		G	GGCTA	С	Т	А	Т	Т	С	Т	G	с
EU106112.1 Spatholobus parviflorus	G C	;	CTATTC	С	с	G	С	А	С	С	А	С	С	А	G	А	G	С	G	ACC		G	GGCTA	С	Т	А	Т	Т	С	Т	G	с
LC080907.1 Spatholobus parviflorus	G C	:	CTATTC	С	С	G	С	А	С	С	А	С	С	А	G	А	G	С	G	ACC		G	GGCTA	С	Т	А	Т	Т	С	Т	-	с
AB924834.1 Spatholobus parviflorus	G C	:	CTATTC	С	с	G	С	А	С	С	А	С	С	А	G	А	G	С	G	ACC		G	GGCTA	с	Т	А	Т	Т	С	Т	G	с
HG004992.1 Spatholobus pulcher	G C	:		Т	А	Т	А	G	Т	Т	А	А	Т	G	G	А	G	Т	G	ACC		G	GGCTA	с	С	А	А	Т	С	с	G	с
HG004955.1 Spatholobus suberectus	тс	:		Т	А	Т	А	G	Т	С	А	А	Т	G	А	А	G	Т	G	ACC		G	GGCTA	с	С	А	А	Т	С	С	G	с
AB925106.1 Spatholobus acuminatus	ТА	. TTT	CTATTC	С	с	Т	с	G	Т	С	С	С	Т	G	G	А	Т	Т	А	ACC		А	GGCTA	с	Т	G	А	G	С	Т	G	А
MG816804.1 Spatholobus littoralis	тс	NNN		С	А	Т	А	G	Т	с	А	А	Т	G	G	Т	G	Т	G	NNN	CCA	N	NNNNN	Т	С	А	А	т	А	С	G	с

Note: *Red: intraspecies variation



The percentage of transitions obtained from the alignment results using the *psbA-trn*H locus was 0.7%, while the percentage of transversions was 2.44% (Table 7). From the alignment using the *mat*K locus, it was determined that the conserved site in the *mat*K mutation was 93.19%, while the variable site was 4.82%. The *mat*K mutation pattern showed a transition percentage of 1.02% while for transversion 0.85% (Table 6).

A high percentage of variable sites in a locus indicates that the locus meets the principle of minimalism, which is an effective means of describing genetic variation between taxa both interspecies and intraspecies. The findings of this study are consistent with previous research, which indicates that *psbA-trn*H has a higher variable site in comparison to *mat*K and rbcL. The results indicate that the psbA-trnH region is an effective locus for identifying both interspecies and intraspecies kinship relationships. The three loci used in this study have a high percentage of conserved sites. A very small percentage of conserved sites represents a weakness of DNA barcoding, as it led to bias in the phylogenetic tree produced (Sumarlina & Napitupulu, 2022).

Phylogenetic trees of the genus *Spatholobus* were constructed with different models. The fittest

model for *rbc*L was K2+G, while the fittest model for matK and psbA-trnH was T92. Construction of a phylogenetic tree in the genus Spatholobus showed clade differences among the markers rbcL, matK, and psbA-trnH (Figure 1). These results showed that *rbc*L, *mat*K, and *psb*A-*trn*H successfully grouped the same species in a separate clade. The phylogenetic tree based on the rbcL marker showed five clades formed, namely S. suberectus, S. parviflorus, S. acuminatus, S. pulcher, and S. gyrocarpus (Figure 1A). The phylogenetic tree based on *mat*K markers also showed five clades formed, namely S. parviflorus, S. suberectus, S. pulcher, S. acuminatus, and S. littoralis (Figure 1C). Meanwhile, the phylogenetic tree based on *psbA-trn*H markers only showed two clades formed, namely S. suberectus and S. pulcher (Figure 1B).

A phylogenetic tree is a diagram that illustrates the relationship of living things based on their evolution. The branches on a phylogenetic tree represent different levels of sequence. The closer the relationship in the phylogenetic tree, the lower the genetic distance between individuals and the higher the genetic similarity. Therefore, closely related individuals will be grouped in the same clade (Anzani et al., 2021).

		Table 5. Mutation pa	ttern of <i>rbc</i> L	
	А	Т	G	С
 А	26,24	$0,22^{(2)}$	$0,22^{(1)}$	$0,00^{(2)}$
Т	$O,OO^{(2)}$	29,25	$O,OO^{(2)}$	0,00 ⁽¹⁾
G	0,00 ⁽¹⁾	$O,OO^{(2)}$	22,15	$0,00^{(2)}$
С	$0,00^{(2)}$	$0,22^{(1)}$	$O,OO^{(2)}$	21,29

⁽¹⁾Transition

(2) Transversion

Ta	ble	6.	Mutation	pattern	of	matK
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	А	Т	G	С
А	30,84	$0,17^{(2)}$	0,17 ⁽¹⁾	$0,17^{(2)}$
Т	$0,00^{(2)}$	39,75	$0, 17^{(2)}$	$0,34^{(1)}$
G	0,17 ⁽¹⁾	0,17	12,51	$0,00^{(2)}$
С	$0,17^{(2)}$	0,34(1)	$0,00^{(2)}$	14,91

(1) Transition

⁽²⁾ Transversion



	А	Т	G	С
А	31,88	$0,69^{(2)}$	0,35 ⁽¹⁾	$0,35^{(2)}$
Т	0,35 ⁽²⁾	45,05	$0,35^{(2)}$	O , OO ⁽¹⁾
G	O , OO ⁽¹⁾	$0,35^{(2)}$	11,78	$0,00^{(2)}$
С	$0,35^{(2)}$	0,35(1)	$0,00^{(2)}$	8,32

⁽¹⁾Transition

(2) Transversion





(C)

Figure 1. Phylogenetic tree of genus Spatholobus. (A) based on rbcL, (B) based on psbA-trnH, and (C), based on *mat*K. The number next to the node shows bootstrap value.





CONCLUSION

The *rbc*L, *mat*K, and *psbA-trn*H regions can be used as markers for DNA barcoding in the genus *Spatholobus* with different specifications. The *rbc*L and *mat*K can be used to distinguish *Spatholobus* at the genus level, while the *psbA-trn*H can be used to distinguish *Spatholobus* at the species level.

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